

CHLAMYDIA TESTING TECHNOLOGIES
(09/2003)

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Reagent Manufacturer(s): Cells – Diagnostic Hybrids; ViroMed, Inc.; American Type Culture Collection (ATCC). **Fluorescent Antibody Reagents** – Wampole Laboratories MicroTrak (Syva), Bio-Rad Laboratories (Kallestad), VWR Scientific Products (Bartels).

Collection Sites: Endocervical, urethral (male and female), nasopharyngeal (infants), conjunctival, vaginal and rectal.

Specimen Handling: Use chlamydia specific transport media, e.g. 2SP, M4, etc. Refrigerate immediately after collection, transport at 2-8⁰ C. If greater than 48 hours from collection to processing, store frozen at -70⁰ C or colder (do not store at -20 C). Specimens can be stored for up to 6 months at -70⁰ C.

Principle of Test: Specimens are inoculated and centrifuged into a medium containing cycloheximide-treated cells (McCoy, BGM, HeLa, etc.). The cells are grown in one-dram shell vials on glass cover slips or in multi-well plates. Specimens are incubated for 48-72 hours and stained with fluorescent labeled antibodies.

Turn-Around Time¹: 2-4 days.

Advantages²: Culture is currently the preferred test for medico legal cases (very high specificity, 99-100%). Culture has been evaluated for several collection sites for which most non-culture methods have not been FDA cleared (approved). Culture preserves the organism for further studies such as “genetic fingerprinting” and antibiotic susceptibility testing.

Disadvantages: The turn around time is from 2-4 days for results (incubation period 48-72 hours). Culture is less sensitive (40-80%) compared to the newer Nucleic Acid Amplified Tests (90-96%). Culture requires expertise in cell culture and is technically more demanding than any of non-amplification tests. The specimen transport and storage times are critical. Culture is very labor intensive and expensive.

Confirmatory Testing: Not Done

Proficiency Testing Programs: Available. Laboratories performing cell culture for chlamydia are required to enroll in HCFA approved proficiency testing programs.

Specimen Adequacy³: Proper specimen collection and transport is critical. Specific training on proper specimen collection and transport requirements is required for all clinicians collecting specimens. Training and periodic retraining should be a part of an ongoing quality assurance program. Laboratories are encouraged to periodically evaluate endocervical specimens for the presence of columnar and cuboidal epithelial cells, provide feedback, and as indicated, training to clinicians.

¹**Turn-Around Time:** is defined as the time the specimen is received in the testing laboratory to the time the specimen is reported out by the testing laboratory within the normal work week (excluding weekends and holidays).

²Only highlights of the advantages and disadvantages are listed. For further information regarding the advantages and disadvantages check the literature and/or with the specific manufacturer.

³Screening Tests to Detect Chlamydia trachomatis and Neisseria gonorrhoeae Infections – 2002 MMWR October 18, 2002/Vol. 51/No. RR-15

Reagent Manufacturers: Wampole Laboratories MicroTrak (Syva), Bio-Rad Laboratories (Kallestad), and others.

FDA Cleared (Approved) Collection Sites¹: Wampole Laboratories MicroTrak (Syva); Female endocervical, male urethral, male urine specimens, and ocular specimens from symptomatic patients. Bio-Rad Laboratories (Kallestad); male urethral and female endocervical specimens.

Specimen Handling (Product Inserts)¹: In General: use the manufacturers specific transport system. Follow the specific manufacturers recommendations for transport conditions and times.

Wampole Laboratories MicroTrak (Syva); Endocervical, urethral and ocular specimens are transported and stored at 2-25⁰C and tested within 7 days of collection. Urine specimens should be transported and stored at 2-8⁰C and tested within 7 days of collection. Do not freeze specimens.

Bio-Rad Laboratories (Kallestad); Specimens are stable at room temperature for up to 7 days, although refrigeration is preferable.

Principle of Test: (Bio-Rad Laboratories (Kallestad)) When treated specimens are added to a microplate well, chlamydia present (antigens) bind to antibody coating the microplate well. Next, an antibody specific for chlamydia is added which binds to chlamydia (antigen) captured by the antibody on the microplate well. A solution of an enzyme-conjugated antibody is added which then binds to the chlamydia antibody. Following a washing step to remove any unbound components in the microplate well an enzyme specific substrate is added to each well. If chlamydia (antigen) is present the substrate reacts with the enzyme and produces a color change that can be detected and measured in a spectrophotometer.

Turn-Around Time²: 1-3 days

Advantages: The Enzyme Immunassay Tests (EIA) are effective for large scale screening (large volume). Viable organisms are not required for the test. EIAs provide a rapid turn around time (3-5 hours) for results. In addition, they are relatively inexpensive and only require moderate technical skill.

Disadvantages: The EIA is less (60-70%) sensitive compared to the Nucleic Acid Amplified Test (90-96%). In addition, the specificity is less than with Cell Culture (specificity 95-99%). The EIA is not FDA cleared (approved) for rectal, respiratory, vaginal, etc. specimens. While the Wampole Laboratories MicroTrak (Syva) EIA test is FDA cleared (approved) for testing urine specimens, the sensitivity and specificity is significantly reduced and screening urines with the EIA is not currently recommended.

Confirmatory Testing: Recommended when screening populations with low prevalence (positivity), e.g. ≤5% positivity. Several methods are available to confirm positive results. A neutralization (blocking assay) and a Direct Fluorescent Antibody test are both routinely used for confirmatory testing of initial positive results.

Proficiency Testing Programs: Available. Laboratories performing EIA for chlamydia are required to enroll in HCFA approved proficiency testing programs.

Specimen Adequacy³ Proper specimen collection and transport is critical. Specific training on proper specimen collection and transport requirements is required for all clinicians collecting specimens. Training and periodic retraining should be a part of an ongoing quality assurance program. Laboratories are encouraged to periodically evaluate endocervical specimens for the presence of columnar and cuboidal epithelial cells, provide feedback, and as indicated, training to clinicians.

¹**Collection Sites and Specimen handling information is directly from product inserts.**

²**Turn-Around Time: is defined as the time the specimen is received in the testing laboratory to the time the specimen is reported out by the testing laboratory within the normal work week (excluding weekends and holidays).**

³**Screening Tests to Detect Chlamydia trachomatis and Neisseria gonorrhoeae Infections – 2002 MMWR October 18, 2002/Vol. 51/No. RR-15**

Reagent Manufacturers: Wampole Laboratories MicroTrak (Syva), Bio-Rad Laboratories (Kallestad), VWR Scientific Products (Bartels).

FDA Cleared (Approved) Collection Sites¹: Wampole Laboratories MicroTrak (Syva); Urethral (males), cervical (females), rectal (symptomatic), conjunctival (symptomatic) and nasopharyngeal specimens (symptomatic). Bio-Rad Laboratories (Kallestad); Urogenital (adult), rectal (adult), ocular (pediatric) and nasopharyngeal (pediatric).

Specimen Handling (Product Inserts)¹: In General: use the manufacturers specific transport system. Follow the specific manufacturers recommendations for transport conditions and times.

Wampole Laboratories MicroTrak (Syva); For best results, store/transport at 20-30⁰C or refrigerated at 2-8⁰C and stain within 7 days of collection. If not stained within 7 days, fixed specimens should be stored at -20⁰C.

Bio-Rad Laboratories (Kallestad); Store the slide in the Pathfinder Collection Kit at room temperature. Process the slide within 7 days.

Principle of Test: The patient specimen is collected, the slide and mailing container are both marked with the patients name and/or ID. The swab is then lightly rolled onto the single wellied-slide provided (within the encircled well area only) and the specimen is fixed immediately (methanol). After the specimen has air dried (2-3 minutes) it is placed in the mailing container and transported to the laboratory. At the laboratory, the slides are then stained with a fluorescent labeled antibody specific for chlamydia. The antibody conjugate binds specifically to any *Chlamydia trachomatis* present in the specimen. A rinse step removes unbound antibody. When slides are viewed under a fluorescence microscope, chlamydia positive specimens contain apple-green elementary bodies contrasted by the red background of the counterstained cells.

Turn-Around Time²: 1-2 days

Advantages: The DFA is FDA cleared (approved) for many different types of specimens. Viable organisms are not required for the test. The test provides a rapid turn around time (1-2 hours) for results. In addition, the DFA is relatively inexpensive, requires moderate technical skill, and the quality of the specimen can be visualized (columnar or cuboidal epithelial cells).

Disadvantages: The DFA is not suitable for large volume testing. The DFA is less (50-70%) sensitive compared to the Nucleic Acid Amplified Test (90-96%). In addition, the specificity is less than with Cell Culture (specificity 95-99%).

Confirmatory Testing: Not Available

Proficiency Testing Programs: Available. Laboratories performing the DFA test for chlamydia are required to enroll in HCFA approved proficiency testing programs.

Specimen Adequacy³ Proper specimen collection and transport is critical. Specific training on proper specimen collection and transport requirements is required for all clinicians collecting specimens. Training and periodic retraining should be a part of an ongoing quality assurance program. Laboratories are encouraged to periodically evaluate endocervical specimens for the presence of columnar and cuboidal epithelial cells, provide feedback, and as indicated, training to clinicians.

¹Collection Sites and Specimen handling information is directly from product inserts.

²**Turn-Around Time:** is defined as the time the specimen is received in the testing laboratory to the time the specimen is reported out by the testing laboratory within the normal work week (excluding weekends and holidays).

³Screening Tests to Detect *Chlamydia trachomatis* and *Neisseria gonorrhoeae* Infections – 2002 MMWR October 18, 2002/Vol. 51/No. RR-15

Reagent Manufacturer(s): Gen-Probe, Inc. (Pace 2, Chlamydia or Gonorrhea, Pace 2C, Combined CT/GC)

FDA Cleared (Approved) Collection Sites¹: Gen-Probe, Inc., Pace 2: Endocervical, male urethral and conjunctival specimens; Pace 2C: Endocervical and male urethral specimens.

Specimen Handling (Product Inserts)¹: **In General:** use the manufacturers specific transport system. Follow the specific manufacturers recommendations for transport conditions and times. Gen-Probe, Inc., Pace 2, Pace 2C: Transport the tubes to the laboratory at 2 to 25⁰C and store at 2 to 25⁰C until tested. Samples should be assayed within 7 days of collection. If longer storage is necessary, process the specimen as described in sample preparation and freeze at -20 to -70⁰C.

Principle of Test: The Gen-Probe Pace system uses a chemiluminescent labeled, single-stranded DNA probe that is complementary to the ribosomal RNA of the target organism (chlamydia). After the ribosomal RNA is released from the organism by heating the labeled DNA probe combines with the chlamydia ribosomal RNA to form a stable DNA:RNA hybrid. The labeled, DNA:RNA hybrid is separated from the non-hybridized probe and is measured in the Gen-Probe luminometer. The test results are calculated as the difference between the response of the specimen and the mean response of the negative reference (control). In the Pace 2C test (combination assay), positive test result must be rerun with CT and GC specific reagents.

Turn-Around Time²: 1-3 days

Advantages: The Gen-Probe Test is effective for large scale screening (large volume). Viable organisms are not required for the test. NAPs provide a rapid turn around time (3-5 hours) for results. In addition, they are relatively inexpensive and only require moderate technical skill. With the Gen-Probe assay, it is also possible to test for chlamydia and gonorrhea from the same specimen.

Disadvantages: The Gen-Probe Test is less sensitive (60-75%) compared to the Nucleic Acid Amplified Test (90-96%). In addition, the specificity is less than with Cell Culture (specificity 95-99%). The NAP is not FDA cleared (approved) for rectal, respiratory, and vaginal specimens.

Confirmatory Testing: Available and recommended when screening populations with low prevalence (positivity), e.g. ≤5% positivity or specimens above the test cut off (CO) to ≤1,500 Relative Light Units (RLU). The Gen-Probe confirmatory test is the Probe Competition Assay (PCA), similar to the EIA neutralization confirmatory procedure.

Proficiency Testing Programs: Available. Laboratories performing NAP tests for chlamydia are required to enroll in HCFA approved proficiency testing programs.

Specimen Adequacy³ Proper specimen collection and transport is critical. Specific training on proper specimen collection and transport requirements is required for all clinicians collecting specimens. Training and periodic retraining should be a part of an ongoing quality assurance program. Laboratories are encouraged to periodically evaluate endocervical specimens for the presence of columnar and cuboidal epithelial cells, provide feedback, and as indicated, training to clinicians.

¹Collection Sites and Specimen handling information is directly from product inserts.

²**Turn-Around Time:** is defined as the time the specimen is received in the testing laboratory to the time the specimen is reported out by the testing laboratory within the normal work week (excluding weekends and holidays).

³Screening Tests to Detect Chlamydia trachomatis and Neisseria gonorrhoeae Infections – 2002 MMWR October 18, 2002/Vol. 51/No. RR-15

Reagent Manufacturer(s): Digene CT/GC Hybrid Capture II Test

FDA Cleared (Approved) Collection Sites¹: Endocervical and male urethral specimens

Specimen Handling (Product Inserts)¹: **In General:** use the manufacturers specific transport system. Follow the specific manufacturers recommendations for transport conditions and times. Specimens may be held for up to two weeks at room temperature and shipped without refrigeration to the testing laboratory. At the testing laboratory, specimens should be stored at 2-8⁰C if the assay is to be performed within one week. If the assay will be performed later than one week, store specimens at -20⁰C for up to 3 months. Specimens collected in Specimen Transport Medium can not be used for culture or other testing methods.

Principle of Test: Specimens are denatured to produce single strands of DNA. A specific single stranded RNA probe is added which recognizes and hybridizes with specific DNA sequences of the CT and GC genomes and plasmids. The resulting RNA/DNA hybrids are then captured on a microtiter plate by antibodies specific to the hybrids. The use of enzyme conjugated antibodies and a substrate produce a signal that is measured on a luminometer. Multiple antibodies, each conjugated to enzyme, bind to each captured hybrid, resulting in a substantial “signal amplification”.

Turn-Around Time²: 1-3 days

Advantages: The Digene CT/GC Hybrid Capture II Test is effective for large scale screening (high volume). Viable organisms are not required for the test. The NAPSA test provides a rapid turn around time (5-6 hours) for results. In addition, it only requires moderate technical skill.

Disadvantages: Insufficient peer reviewed publications are available for the Digene CT/GC Hybrid Capture II Test to accurately predict test sensitivity and specificity. However, based on the few preliminary publications and the signal amplification technology, the sensitivity appears to be slightly higher than the current Gen-Probe Pace 2/2C Test and less than the Nucleic Acid Amplified Tests (NAAT). Test specificity appears to compare with the Gen-Probe Pace Test. The NAPSA is not FDA cleared (approved) for rectal, respiratory, and vaginal specimens.

Confirmatory Testing: Not Available

Proficiency Testing Programs: Available. Laboratories performing NAPSA for chlamydia are required to enroll in HCFA approved proficiency testing programs.

Specimen Adequacy³ Proper specimen collection and transport is critical. Specific training on proper specimen collection and transport requirements is required for all clinicians collecting specimens. Training and periodic retraining should be a part of an ongoing quality assurance program. Laboratories are encouraged to periodically evaluate endocervical specimens for the presence of columnar and cuboidal epithelial cells, provide feedback, and as indicated, training to clinicians.

¹Collection Sites and Specimen handling information is directly from product inserts.

²**Turn-Around Time:** is defined as the time the specimen is received in the testing laboratory to the time the specimen is reported out by the testing laboratory within the normal work week (excluding weekends and holidays).

³Screening Tests to Detect Chlamydia trachomatis and Neisseria gonorrhoeae Infections – 2002 MMWR October 18, 2002/Vol. 51/No. RR-15

NUCLEIC ACID AMPLIFICATION TESTS

GENERAL COMMENTS:

ADVANTAGES:

Nucleic Acid Amplification Tests (NAATs) are the most sensitive and specific tests currently available for the detection of chlamydia. The sensitivity of NAATs are in the mid to high 90s, with specificity equal to cell culture (99-100%). While culture is the preferred test for medico legal cases, most experts have suggested that NAATs for chlamydia could be used as an alternative to culture if culture is not available. If a NAAT is used for medico legal purposes, it is currently recommended that an alternate NAAT be performed on all initial positive results^{1,2}. Viable organisms are not required for the test. NAATs provide a rapid turn around time (3-5 hours) for results

DISADVANTAGES:

The NAATs are technically demanding and special areas³ within laboratories need to be utilized in order to run these tests due the increased potential for cross contamination of specimens. The specimen transport and storage times with most NAATs are critical (excluding Gen-Probe APTIMA Combo 2). The NAATs are not FDA cleared (approved) for respiratory, conjunctival, vaginal or rectal specimens. The NAATs are still fairly expensive, running from 2-4 times the diagnostic reagent costs for Non-Amplified Tests. No confirmatory or supplemental tests are currently FDA cleared for the NAATs. However, Gen-Probe currently provides Analyte Specific Reagents⁴ (ASR) for *Chlamydia trachomatis* and *Neisseria gonorrhoeae* that may be used to “confirm” positive specimens. Laboratories initiating any Nucleic Acid Amplified Test (NAAT) should repeat all positive specimens during their initial verification process and evaluate the need to continue to “confirm” positive specimens on a routine basis. For additional information on “confirmation” of positive specimens refer to CDCs, “Screening Tests To Detect *Chlamydia trachomatis* and *Neisseria gonorrhoeae* Infections – 2002”, MMWR, October 18, 2002/Vol. 51/No. RR-15².

¹CDC. Sexually Transmitted Diseases Treatment Guidelines. 2002. MMWR 2002;51:72-73.

²CDC. Screening Tests to Detect *Chlamydia trachomatis* and *Neisseria gonorrhoeae* Infections. 2002. MMWR 2002;51:23.

³Each manufacturer has specific laboratory facility and space requirements for their specific NAAT technology. Any laboratories initiating NAATs should check with the manufacturer on the facility and space requirements before making any test methodology decisions.

⁴See Appendix A for more information on Analyte Specific Reagents (ASR).

Reagent Manufacturer: Roche Diagnostics; Roche (COBAS AMPLICOR CT/NG Tests) and Roche (MICROTITER AMPLICOR CT/NG Test)

FDA Cleared (Approved) Collection Sites¹: Urine from males and females, endocervical swab specimens, male urethral swab specimens (symptomatic or asymptomatic).

Specimen Handling (Product Inserts)¹: Urine specimens may be transported to the test site at 18–25⁰C. Urine specimens are stable for 24 hours at 18–25⁰C. Urine specimens that require shipment to off-site test centers must be shipped via overnight delivery with guaranteed arrival within 24 hours; shipment can be at 18–25⁰C. If urine specimens are shipped at 18–25⁰C, they should be stored at 2 – 8⁰C until time of shipment to ensure that the period of 18–25⁰C storage does not exceed 24 hours. Urine specimens that will not be processed within 24 hours of collection must be stored at 2–8⁰C and must be processed within 7 days of collection. Urine specimens that cannot be processed within 7 days of collection can be stored at –20⁰C or lower for up to 2 months.

Swab specimens may be transported to the test site at 18–25⁰C provided that the total time of storage and transport at 18–25⁰C is less than 1 hour. Refrigerate swab specimens if transport to the laboratory is delayed for more than one hour from the time of collection. If used for culture, specimens should be handled using recommendations for culture (-70⁰C if not cultured within 24 hr). Swab specimens that require off-site laboratories should be shipped at refrigerated temperature as soon as possible after collection according to the laboratory's procedures for the transport of chlamydial culture specimens. Store swab specimens that are not tested upon receipt at the testing laboratory within 7 days of collection must be stored at –20⁰C or colder and tested within 30 days of collection.

Principle of Test: Specimens are treated to release the chlamydia DNA and make it accessible for replication and detection. After treatment two short, labeled (biotinylated) nucleotides (primers) that are complementary to the chlamydia target DNA along with an enzyme (polymerase) and excess nucleotides are added to the reaction mixture. The primers bind to the specific chlamydia target DNA. The polymerase enzyme extends each DNA sequence utilizing the excess nucleotides to form a complementary DNA sequence (amplicon). This amplification process occurs when the reaction mixture is incubated in a DNA thermal cycler and alternately heated and cooled, multiple times (cycles). Each cycle exponentially increases the amount of target DNA. After amplification, the amplicons are hybridized by utilizing a specific DNA probe that is complementary to the amplicons. This probe is bound to a solid support (microparticles) and used to capture the amplicons. The last step in this process is the assay detection system that uses a standard enzyme/substrate reaction in order to detect color change and subsequent positive specimens.

Turn-Around Time²: 1-3 days

Advantages: The PCR NAAT is FDA cleared for both chlamydia and gonorrhea screening on the same specimen. In addition, the test is FDA cleared (approved) for urine specimens (sensitivity for urine specimens is slightly lower compared to swab specimens). The Roche COBAS AMPLICOR instrument is semi-automated. The Roche PCR test comes in two configurations, the COBAS AMPLICOR system for the smaller volume laboratories and the Roche Microtiter Plate test for larger volume laboratories. The PCR test has the ability to include an internal amplification control that enables the laboratory to determine the presence of inhibitors that may produce a false negative result. The PCR test has the ability to prevent false positive results (contamination) from any previously amplified products (amplicons) with AmpErase.

Disadvantages: Only limited number of specimens can be run with the Roche COBAS AMPLICOR system (similar to the Abbott LCx instrument).

Proficiency Testing Programs: Available. Laboratories performing NAATs for chlamydia are required to enroll in HCFA approved proficiency testing programs.

Specimen Adequacy³ Proper specimen collection and transport is critical. Specific training on proper specimen collection and transport requirements is required for all clinicians collecting specimens. Training and periodic retraining should be a part of an ongoing quality assurance program. Laboratories are encouraged to periodically evaluate endocervical specimens for the presence of columnar and cuboidal epithelial cells, provide feedback, and as indicated, training to clinicians.

¹Collection Sites and Specimen handling information is directly from product inserts.

²Turn-Around Time: is defined as the time the specimen is received in the testing laboratory to the time the specimen is reported out by the testing laboratory within the normal work week (excluding weekends and holidays).

³Screening Tests to Detect Chlamydia trachomatis and Neisseria gonorrhoeae Infections – 2002 MMWR October 18, 2002/Vol. 51/No. RR-15

TRANSCRIPTION MEDIATED AMPLIFICATION (TMA) TEST

Reagent Manufacturer: Gen-Probe, Incorporated

FDA Cleared (Approved) Collection Sites¹: Endocervical and male urethral swab specimens and in female and male urine specimens.

Specimen Handling (Product Inserts)¹: Transport swab specimens to the laboratory and store at 2 to 25C until tested. Specimens should be assayed with the Gen-Probe Amplified Chlamydia Trachomatis Assay within 7 days of collection. If longer storage is needed, freeze at -20 to -70C.

Transport urine specimens to the laboratory at ambient temperature within 24 hours of collection, or at 2 to 8C within 7 days of collection. Store urines at 2 to 8C within 7 days of collection. Urine specimens should be assayed with the Gen-Probe Amplified Chlamydia Trachomatis Assay within 7 days of collection. If longer storage is needed, mix the specimen, then aliquot into single-use portions and freeze at -70C. Once thawed, do not re-freeze urines.

Principle of Test: Specimens are processed (treated) to release chlamydia specific RNA and amplified utilizing Transcription-Mediated Amplification (TMA). The TMA system uses specific ribosomal RNA target via DNA intermediates. After amplification the amplified products (amplicons) are detected by nucleic acid hybridization (probe). The chemiluminescent single-stranded DNA probe is complementary to the amplicons. The labeled DNA probe combines with the amplicons to form a stable RNA:DNA hybrid. The labeled RNA:DNA hybrids are measured in a luminometer. A positive result is a luminometer reading greater than or equal to the cut-off value. A value less than the cut-off is a negative result.

Turn-Around Time²: 1-3 days

Advantages: The TMA is FDA cleared (approved) for urine specimens (sensitivity for urine specimens is slightly lower compared to swab specimens). Because TMA is isothermal, no thermocycling equipment is needed. Since the amplification target is RNA and RNA is more labile than DNA the potential for cross contamination is reduced.

Disadvantages: The TMA NAAT is not FDA cleared for gonorrhea screening. The current configuration of the TMA requires multiple pipeting steps and is labor intensive. A large volume version is available which uses a semi-automated Tecan pipetting/diluting station.

Proficiency Testing Programs: Available. Laboratories performing NAATs for chlamydia are required to enroll in HCFA approved proficiency testing programs.

Specimen Adequacy³ Proper specimen collection and transport is critical. Specific training on proper specimen collection and transport requirements is required for all clinicians collecting specimens. Training and periodic retraining should be a part of an ongoing quality assurance program. Laboratories are encouraged to periodically evaluate endocervical specimens for the presence of columnar and cuboidal epithelial cells, provide feedback, and as indicated, training to clinicians.

¹Collection Sites and Specimen handling information is directly from product inserts.

²**Turn-Around Time:** is defined as the time the specimen is received in the testing laboratory to the time the specimen is reported out by the testing laboratory within the normal work week (excluding weekends and holidays).

³Screening Tests to Detect Chlamydia trachomatis and Neisseria gonorrhoeae Infections – 2002 MMWR October 18, 2002/Vol. 51/No. RR-15

Reagent Manufacturer: Gen-Probe, Incorporated

FDA Cleared (Approved) Collection Sites¹: Endocervical and male urethral swab specimens and in female and male urine specimens. The assay may be used to test specimens from symptomatic and asymptomatic individuals.

Specimen Handling (Product Inserts)¹: Swab specimens must be transported to the laboratory in the swab specimen transport medium and tube. Swab specimens must be transported to the laboratory at 2^o to 30^oC and tested within 60 days of collection.

Urine specimens can be transported to the laboratory at 2^o to 30^oC in either the primary collection device (urine cup) or in the urine specimen transport tube. Urine specimens must be transferred into the GEN-PROBE specimen transport tube within 24 hours of collection and before being assayed. After transfer, urine specimens can be stored at 2^o to 30^oC for up to 30 days after collection.

The APTIMA Combo 2 Assay Unisex Swab Specimen Collection Kit for Endocervical and Urethral Swab Specimens and the APTIMA Combo 2 Assay Urine Specimens Collection Kit for Male and Female Urine Specimens are intended to be used only with the GEN-PROBE APTIMA Combo 2 Assay. Performance has not been established with other products.

Principle of Test: The GEN-PROBE APTIMA Combo 2 Assay combines the technologies of target capture, Transcription-Mediated Amplification (TMA) and Dual Kinetic Assay (DKA).

Target Capture uses capture oligonucleotides and magnetic microparticles to capture target nucleic acid and separate potentially inhibiting substances from samples, thereby isolating the desired nucleic acid targets for amplification.

Transcription-Mediated Amplification (TMA) uses specific ribosomal RNA target via DNA intermediates to potentially produce billions of copies of target nucleic acid.

The Dual Kinetic Assay (DKA) uses single stranded chemiluminescent DNA probes that are complementary to a region of each target amplicon and are labeled with different acridinium ester molecules. The labeled probe combines with amplicon to form stable RNA:DNA hybrids. The labeled RNA:DNA hybrids are measured in a luminometer. In DKA, differences in the kinetic profiles of the *C. trachomatis* and *N. gonorrhoeae* labeled probes allow for the differentiation of signal; kinetic profiles are derived from measurements of photon output during the detection read time. The chemiluminescent detection reaction for *C. trachomatis* signal has very rapid kinetics and has the “flasher” kinetic type. The chemiluminescent detection signal for *N. gonorrhoeae* is relatively slower and has the “glower” kinetic type. Assay results are determined by a cut-off based on the total Relative Light Unit (RLU) and the kinetic curve type.

Turn-Around Time²: 1-3 days

Advantages: The APTIMA Combo 2 is FDA cleared (approved) for urine specimens (sensitivity for urine specimens is equal to or slightly higher than for swab specimens). Because TMA is isothermal, no thermocycling equipment is needed. Since the amplification target is RNA and RNA is more labile than DNA the potential for cross contamination is reduced. The utilization of Target Capture should reduce or eliminate potential inhibitors. The Dual Kinetic Assay allows simultaneous detection of chlamydia and gonorrhea. Gen-Probe has recently made Analyte Specific Reagents (ASR) available for confirmatory testing. The availability of single ASR should reduce the price of confirmatory testing or screening for individual agents.

Disadvantages: The APTIMA Combo 2 is a combined test for CT and GC. It is generally not economical, practical or ethical to attempt to report only single results.

Proficiency Testing Programs: Available. Laboratories performing NAATs for chlamydia are required to enroll in HCFA approved proficiency testing programs.

Specimen Adequacy³ Proper specimen collection and transport is critical. Specific training on proper specimen collection and transport requirements is required for all clinicians collecting specimens. Training and periodic retraining should be a part of an ongoing quality assurance program. Laboratories are encouraged to periodically evaluate endocervical specimens for the presence of columnar and cuboidal epithelial cells, provide feedback, and as indicated, training to clinicians.

¹Collection Sites and Specimen handling information is directly from product inserts.

²Turn-Around Time: is defined as the time the specimen is received in the testing laboratory to the time the specimen is reported out by the testing laboratory within the normal work week (excluding weekends and holidays).

³Screening Tests to Detect Chlamydia trachomatis and Neisseria gonorrhoeae Infections – 2002 MMWR October 18, 2002/Vol. 51/No. RR-15

Reagent Manufacturer: Becton Dickinson (BD ProbeTec ET System)

FDA Cleared (Approved) Collection Sites¹: The BDProbeTec ET System is designed to detect the presence of Chlamydia trachomatis in endocervical swabs, male urethral swabs and male and female urine specimens using the appropriate collection method. Specimens may be from symptomatic or asymptomatic females for the BDProbeTec ET CT and GC Assays, from symptomatic or asymptomatic males for the BDProbeTec ET CT Assay, and from symptomatic males for the BDProbeTec ET GC Assay.

Specimen Handling (Product Inserts)¹: The CULTURETTE DIRECT collection swab and Mini-Tip CULTURETTE DIRECT swab must be stored and transported to the laboratory and/or test site at 2 – 27⁰C within 4-6 days of collection. Storage up to 4 days has been validated with clinical specimens; storage up to 6 days has been demonstrated with seeded specimens. NOTE: if specimens cannot be transported directly to the testing laboratory under ambient temperatures (15 – 27⁰C) and must be shipped, an insulated container with ice should be used with either overnight or 2-day delivery vendor. BD has FDA clearance for their “wet swab” transport system. In the “wet swab” transport system the swab dilution medium has been added to the transport tube. In addition, BD has received FDA clearance for their BD Viper specimen processing station.

If the test site adds the UPP², store and transport urine specimens to the test site at 2 – 8⁰C within 4-6 days of collection. Do not freeze the specimen.

If the UPP is added at the collection site, store and transport urine specimens to the laboratory or test site at 2 – 8⁰C within 4-6 days of collection or at 15 – 27⁰C within 2 days of collection. Do not freeze the specimen.

Principle of Test: The SDA is an amplified DNA assay based on the simultaneous amplification and detection of target DNA using amplification primers and a fluorescent labeled detector probe. The SDA reagents are dried in two separate disposable microwell strips. The processed sample is added to the priming microwell that contains the amplification primers, fluorescent labeled detector probe, and other reagents necessary for amplification. After incubation, the reaction mixture is transferred to the amplification microwell, which contains two enzymes (a DNA polymerase and a restriction endonuclease) necessary for the SDA. The amplification microwells are sealed to prevent contamination and then incubated in a thermally controlled fluorescent reader that monitors each reaction for the generation of amplified products. The presence or absence of chlamydia is determined by comparing specimen results to a pre-determined cutoff value. A positive result has a reading greater than or equal to the cut-off value. A value less than the cut-off is a negative result.

Turn-Around Time³: 1-3 days

Advantages: The SDA is FDA cleared (approved) for urine specimens (sensitivity for urine specimens is slightly lower compared to swab specimens). The SDA offers an optional amplification control to monitor each specimen for the presence of an inhibitor that may produce a false negative result. The SDA has a high throughput, up to 564 patient specimens (with no amplification control, chlamydia only) per 8 hour shift. The new “wet swab” transport effectively reduces the specimen processing time and may reduce the cost of labor.

Disadvantages: The SDA has been reported to have some problems with cross contamination from the internal control. In addition several papers have indicated that the SDA GC test may cross react with other Neisseria strains. The new “wet swab” transport is more expensive than the previous swab transport system and may add to the diagnostic cost of the test.

Proficiency Testing Programs: Available. Laboratories performing NAATs for chlamydia are required to enroll in HCFA approved proficiency testing programs.

Specimen Adequacy⁴ Proper specimen collection and transport is critical. Specific training on proper specimen collection and transport requirements is required for all clinicians collecting specimens. Training and periodic retraining should be a part of an ongoing quality assurance program. Laboratories are encouraged to periodically evaluate endocervical specimens for the presence of columnar and cuboidal epithelial cells, provide feedback, and as indicated, training to clinicians.

¹Collection Sites and Specimen handling information is directly from product inserts.

²UPP-Urine Processing Pouch

³Turn-Around Time: is defined as the time the specimen is received in the testing laboratory to the time the specimen is reported out by the testing laboratory within the normal work week (excluding weekends and holidays).

⁴Screening Tests to Detect Chlamydia trachomatis and Neisseria gonorrhoeae Infections – 2002 MMWR October 18, 2002/Vol. 51/No. RR-15

Appendix A

Analyte Specific Reagents

The Analyte Specific Reagents regulation rule was issued in the Federal Register November 21, 1997, Volume 62, Number 225. The rule essentially allowed certain individual reagents to be available for laboratories to use in their own in-house tests. They do not require manufacturer's to submit a 510(k) or PMA.

The APTIMA CT and GC Analyte Specific Reagent (ASR) products can be used for the detection of *Chlamydia trachomatis* or *Neisseria gonorrhoeae* rRNA. The manufacturer may make no performance, stability, or specimen claims for these products. Reagents are specific for an analyte(s) and are not sold as a "kit". These reagents are not FDA cleared and the customer must verify their performance. For additional information on test verification and validation see the Federal Register January 24, 2003, Volume 68, Number 16, Section 493.1253.

The manufacturer is not allowed to troubleshoot any possible performance related problems with or for the testing laboratory. The manufacturer may refer a laboratory that needs assistance to another laboratory running the assay. The manufacturer isn't allowed to provide any procedures to the testing laboratory. In addition, the manufacturer cannot solicit customers for these products. However, they can respond to unsolicited requests for information from laboratories or physicians.

The testing laboratory must include a disclaimer statement when reporting results as follows: "This test was developed and its performance characteristics determined by (Specific Laboratory Name). It has not been cleared or approved by the U.S. Food and Drug Administration."

In addition to a disclaimer statement, results should be reported consistent with the CDC Screening Guidelines¹, "When additional testing has been performed the laboratory should report the results of both the screening test and the additional tests, as well as the overall interpretation."

¹Screening Tests to Detect *Chlamydia trachomatis* and *Neisseria gonorrhoeae* Infections – 2002 MMWR October 18, 2002/Vol. 51/No. RR-15